



Anisakid Larvae from Anchovies in the South Coast of Korea

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Abstract: Anisakiasis (anisakidosis) refers to a foodborne zoonosis caused by ingesting raw or undercooked marine fish or cephalopods infected with anisakid larvae. The present study was performed to investigate the prevalence of anisakid larvae in anchovies (*Engraulis japonica*) purchased from 2 local markets in Gyeongsangnam-do, the Republic of Korea (= Korea), during 2018-2019. Anchovies were transported to our laboratory and examined by pepsin-HCl artificial digestion technique followed by microscopic observations and molecular analyses. The overall prevalence of anisakid larvae was 19.5% (39/200), from which a total of 51 larvae (av. 1.3 larvae/infected anchovy) were recovered. Sequencing of the larvae targeting the ITS region, including ITS1, 5.8S rRNA, and ITS2 genes confirmed the species of larvae as *Anisakis pegreffii* (54.9%; 28/51), *Hysterothylacium sinense* (23.5%; 12/51), and *Hysterothylacium aduncum* (21.5%; 11/51). The results suggested that anchovies could be a potential source of human anisakiasis in Korea.

Key words: *Anisakis pegreffii*, *Hysterothylacium sinense*, *Hysterothylacium aduncum*, anisakid larva, anchovy, Korea

Anisakiasis refers to a foodborne zoonosis caused by infection of people with larval stages of anisakid nematodes belonging to the families Anisakidae or Raphidascarididae, including the genera *Anisakis*, *Pseudoterranova*, and *Contracaecum* [1]. The disease may occur when people ingest raw marine fish or cephalopods, which contain third-stage larvae in their viscera or muscle. The first reported anisakiasis was 11 cases of previously healthy adults who experienced violent abdominal colic after ingestion of marine fish in the Netherlands [2]; larval anisakid nematodes were found deeply buried in their gut wall. After then, anisakiasis has been reported from almost all over the world. Over 90% of the total anisakiasis cases (about 20,000) were reported from Japan, followed by the Netherlands, Germany, France, Spain, and the Republic of Korea (= Korea) [1]. The clinical symptoms of invasive anisakiasis include severe abdominal pain, nausea, and vomiting, and the larvae can be removed at gastrointestinal endoscopy or by surgery [1]. In some cases of invasive anisakiasis, the larvae rarely

penetrate other sites such as the throat or visceral organs [1,3]. To prevent this infection, marine fish needs to be immediately eviscerated after fishery and be cooked at temperatures greater than 60°C or frozen at -20°C for at least 24 hr prior to consumption [1].

People in Korea like to consume raw or lightly cooked or salted fish. Consequently, since Kim et al. [3] recorded the first case of human anisakiasis in Korea, a total of 645 anisakiasis cases have been reported until 2015 [4]. Among 203 clinical cases, the majority (165 cases; 81.3%) was due to *Anisakis* type I larvae [4]. Based on morphological analyses, these *Anisakis* type I larvae were diagnosed as *Anisakis simplex* [5-7]. However, 25 out of 26 larvae (96.2%) extracted from human patients during 2000-2013 in Korea were confirmed to be *Anisakis pegreffii* using molecular analyses [8]. More recently, all 20 larvae (100%) extracted from patients by endoscopy in a health check-up center in 2017-2019 were also molecularly confirmed to be *A. pegreffii* [9]. The larvae of *A. simplex* and *A. pegreffii* are morphologically indistinguishable (*Anisakis* type I), and therefore molecular analyses were required to determine the precise species [10].

Several studies have been performed on the prevalence of anisakid larvae in various marine fishes in Korea [11-15]. However, the infection status of anchovies (*Engraulis japonica*) has

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seldom been reported except in a study by Song et al. [11], which used the morphological method for species identification. In the present study, we investigated the infection status of anchovies purchased from 2 markets in the south coast of Korea with anisakid larvae using both morphological and molecular methods. The internal transcribed spacer (ITS) region, including the ITS1, 5.8S rRNA, and ITS2 genes, were sequenced for molecular studies.

A total of 200 anchovies (100 from each area) were purchased from 2 local fish markets located in a middle (area A) and a western part (area B) of Gyeongsangnam-do, Korea (50 km apart from each other). The area A was surveyed in October 2018 and the area B was studied in May 2019. The anchovies were stated to have been caught near the coast of the South Sea, Korea. In total, 51 anisakid larvae were collected from 200 anchovies, and all the larvae collected were molecularly analyzed.

To isolate the larvae from anchovies, the pepsin-HCl artificial digestion method was used. The procedure was as it follows. Identification of the fish species and measurement of the size and weight of each fish. Grinding the fish in a mortar with

pestle. Transferring the flesh of the fish into a beaker and pour artificial gastric juice (8 ml HCl + 6 g pepsin 1: 10,000 + 1,000 ml of 0.85% saline). Mixing the mixture well and keeping it at 37°C for 2 hr with occasional stirring. Filtering it through a 1 × 1 mm mesh. Adding 0.85% saline and letting it stand until sediment came down. Discarding the supernatant carefully not to throw away the sediments. Repeating the washes until the supernatant became clear. Transferring the sediment into a Petri dish and examination under a stereomicroscope.

Anisakid larvae collected from the sediment were washed in water, cleared in lactophenol solution, and observed under an optical microscope (Leica D2500, Leica Microsystems, Wetzlar, Germany). Morphologically, third-stage larvae of *Anisakis* type I and 2 *Hysterothylacium* spp. (*H. sinense* and *H. aduncum*) were identified.

In order to obtain a molecular (species-specific) diagnosis, the total genomic DNA (gDNA) was extracted using the DNeasy Blood and Tissue kit (Qiagen, Hilden, Germany) according to the manufacturer's instructions. Molecular identification was carried out by sequencing of the ITS (ITS1, 5.8 subunit rRNA, and ITS2) genes. Amplification of the nuclear ribosomal region was performed using the ITS A (5'-GTC GAA TTC GTA GGT GAA CCT GCG GAA GGA TCA-3') and ITS B (5'-GCC GGA TCC GAA TCC TGG TTA GIT TCT TTT CCT-3') [16] with 2X MasterMix (MGmed, Seoul, Korea), containing 10 pmol of each primer. The mixture was undergone initial denaturation at 95°C for 10 min, followed by 35 cycles of denaturation at 95°C for 30 sec, annealing at 55°C for 30 sec, and extension at 72°C for 80 sec with a final extension at 72°C for 5 min. Automated DNA sequencing experiments on ITS PCR products were performed at Macrogen Co. (Seoul, Korea).

Anisakid larvae were detected in 39 (19.5%) out of 200 anchovies examined (Fig. 1A, B), and a total of 51 larvae were collected (av. 1.3 larvae/infected anchovy) (Table 1). The larvae were divided into 3 different morphological types. Larvae of *Anisakis* type I (*A. pegreffii*) (Group 1) had a boring tooth at its anterior end, a long ventriculus, and a mucron at its caudal



Fig. 1. Anchovy purchased from a local market in Gyeongsangnam-do. (A) Whole fish. (B) Sashimi of the anchovy. Scale bar = 1 cm.

Table 1. Infection status with anisakid larvae in anchovies collected from the south coast of Korea

Area in Gyeongsangnam-do	Length of anchovy (cm)	Weight of anchovy (g)	No. of anchovies examined	No. of infected (%)	No. of larvae isolated (density per anchovy)
A (middle part)	11.1 (6.7-12.5)	8.8 (5.6-13.5)	100	25 (25.0)	32 (1.3)
B (western part)	10.4 (9.5-11.5)	7.1 (5.3-10.6)	100	14 (14.0)	19 (1.4)
Total	10.7	9.9	200	39 (19.5)	51 ^a (1.3)

^aThese consisted of 28 (54.9%) *Anisakis pegreffii*, 12 (23.5%) *Hysterothylacium sinense*, and 11 (21.5%) *Hysterothylacium aduncum*, which were confirmed by molecular analysis targeting the ITS region.

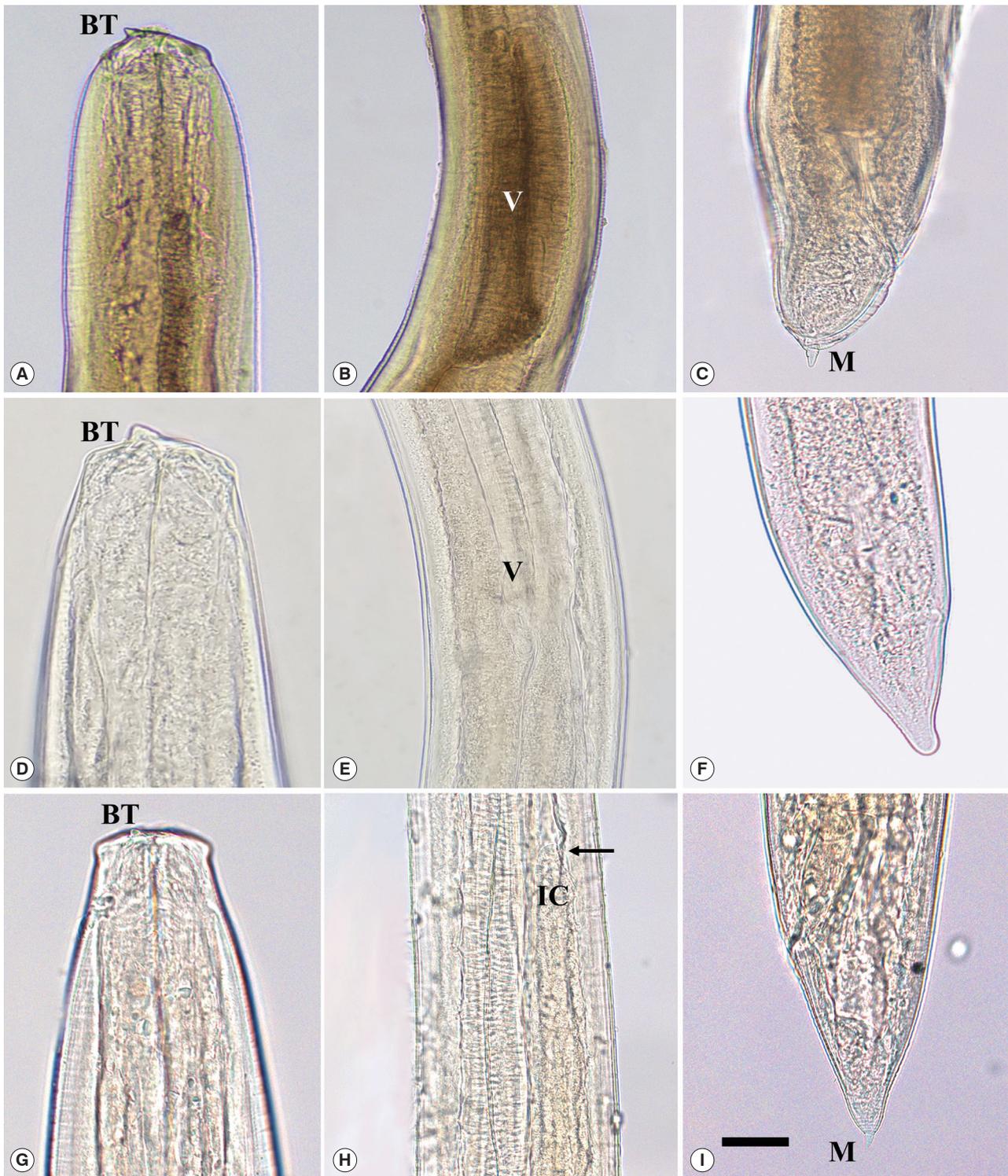


Fig. 2. Microscopic images of third-stage larvae of *Anisakis pegreffii* (A-C), *Hysterothylacium sinense* (D-F), and *Hysterothylacium aduncum* (G-I). Scale bar=0.1 mm. (A) Anterior extremity of *A. pegreffii* larva showing a boring tooth (BT). (B) Middle part showing a long ventriculus (V). (C) Posterior extremity with a mucron (M). (D) Anterior end of *Hysterothylacium sinense* larva with a boring tooth (BT). (E) Middle part showing a small ventriculus (V). (F) Posterior extremity showing its caudal end without a mucron. (G) Anterior extremity of *Hysterothylacium aduncum* larva with a small boring tooth (BT). (H) Middle part showing a long intestinal cecum (IC; arrow indicates its terminal portion). (I) Posterior extremity showing its caudal end with a mucron (M).

end (Fig. 2A-C). Larvae of *Hysterothylacium sinense* (Group 2) had a large boring tooth at its anterior end, a small ventriculus with a long intestinal cecum and a long ventricular appendage, but not a structure like a mucron at its caudal end (Fig. 2D-F). Larvae of *Hysterothylacium aduncum* (Group 3) had a small boring tooth at its anterior end, a small ventriculus with a long intestinal cecum, and a long ventricular appendage, and a mucron at its posterior end (Fig. 2G-I).

The species-specific identification of the anisakid larvae was supported by sequencing of the ITS region (Fig. 3). Group 1 larvae (*Anisakis* type I; 28/51, 54.9%) showed 100% identity with *A. pegreffii* (AB196671) and 99.5% with *A. simplex* (AB277822). Group 2 larvae (12/51, 23.5%) showed 100% identity with *H. sinense* (KX817295), 79.9% with *H. aduncum* (KX110074), and 60.3% with *A. pegreffii* (AB196671). Group 3 larvae (11/51, 21.5%) showed 100% identity with *H. aduncum* (KX110074), 79.9% with *H. sinense* (KX817295), and 63.4% with *A. pegreffii* (AB196671). The prevalence of anisakid larvae in anchovies purchased from 2 surveyed areas was a little different from each other; it was higher in area A (25/100; 25.0%) than in area B (14/100; 14.0%) (Table 1).

The economic condition of Korea has developed remarkably over several decades, and the trends of parasite diseases have changed noticeably. Nowadays, foodborne parasites, in-

cluding liver and intestinal flukes and anisakid larvae, have become the major groups of epidemiological significance, whereas, in the past, soil-transmitted helminthiases were ranked as the most prevalent parasitic diseases [1].

A. pegreffii was originally reported from a Mediterranean monk seal in 1955 by Campana-Rouget and Biocca [17]. The third-stage larvae of *A. pegreffii* are found in various species of marine fish and cephalopods and morphologically classified as *Anisakis* type I, the same type as *A. simplex*. Since the larvae of *A. pegreffii* and *A. simplex* are almost indistinguishable each other, molecular methods, including PCR-RFLP and nucleotide sequencing, were applied for species identification [18-23]. The main geographic habitat of *A. pegreffii* was reported to be the Mediterranean Sea and the Southern Atlantic Ocean [24]. The first case of human anisakiasis caused by *A. pegreffii* was reported in Italy [18]. Later, studies using molecular methods have found that this species also occurs frequently in Korea and Japan [15]. Subsequently, human infections have been found in Japan, Italy, and Korea [8,9,19-22]. In Korea, a total of 35 *A. pegreffii* infections have been molecularly diagnosed, including 15 cases in 2015 [8] and 20 cases in 2019 [9].

In our study, 3 species of anisakid larvae were isolated from anchovies caught from the south coast of Korea. Among 51 larvae isolated, 54.9% (28/51) were molecularly confirmed to

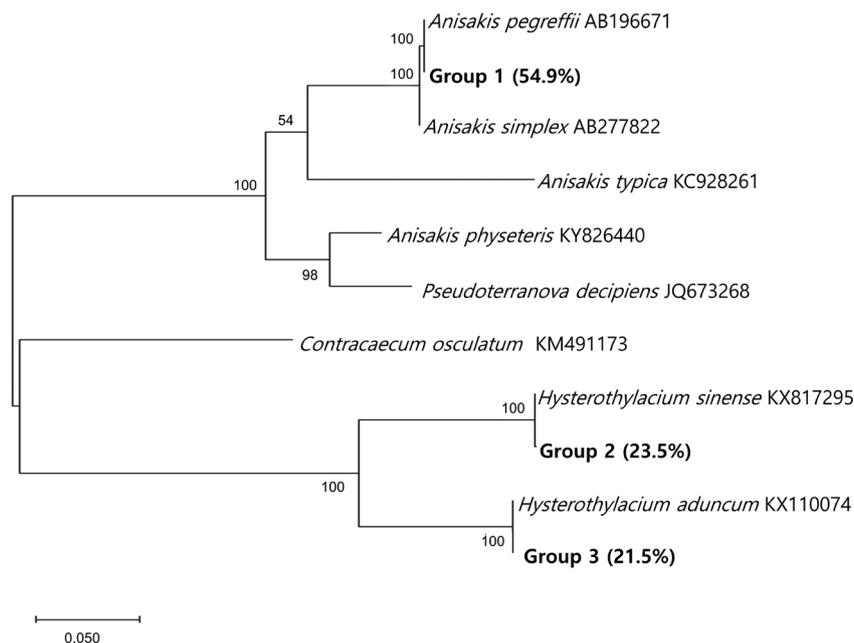


Fig. 3. A phylogenetic tree of anisakid nematodes based on sequences of the ITS region. This tree was constructed from a representative selection of sequences available in GenBank using the maximum-likelihood method employing Tamura-nei model of nucleotide substitution with 1,000 bootstrap replications. Scale bars indicate nucleotide substitutions per site.

be *A. pegreffii*. According to Sohn et al. [4], 1.3% of anisakiasis patients (4/311) reported in Korea recalled that they had eaten anchovies. Therefore, anchovies are suspected as one of the potential sources of human anisakiasis in Korea.

The genus *Hysterothylacium* Ward and Magath, 1917 (Anisakidae) comprises of more than 60 species [25]. The larval stages of these worms mostly parasitize the viscera and tissues of various fish and invertebrate hosts which act as paratenic or intermediate hosts [26,27]. The adult worms of *Hysterothylacium sinense* Li, An and Zhang, 2007 were discovered from the intestine of marine fishes caught off the Yellow Sea, China [25], and their third-stage larvae were found from the cinnamon flounder (*Pseudorhombus cinnamomeus*) caught from the Yellow Sea, China [28]. The presence of adult *Hysterothylacium aduncum* (Rudolphi, 1802) Deardorff and Overstreet, 1981 was reported mainly in temperate and cold waters, including the North-Eastern Atlantic, Seas of the North of Europe, the Mediterranean Sea, Adriatic Sea, Pacific and Atlantic waters of North America, waters of Australia, and the southwest Atlantic Ocean [29]. Various species of fish caught from the south coast of Korea, including the sea eels (*Astroconger myriaster*) and chub mackerel (*Scomber japonicus*), were reported to be infected with *H. aduncum* larvae [9,12-14]. Our study first demonstrated the presence of *H. aduncum* and *H. sinense* larvae in anchovies using molecular analysis.

Among the species of *Hysterothylacium*, human infections have been known only in *H. aduncum*; 2 cases from Japan and Spain [30,31]. In these reports, the specific identification of the larvae was performed only by morphological methods [30,31]. Thus, further (molecular) studies are required to determine whether *H. aduncum* can truly infect humans. In addition, the possibility of human infection with *H. sinense* also needs to be elucidated.

The prevalence of anisakid larvae in anchovies purchased from 2 different areas (middle and western parts of Gyeong-sangnam-do) was different by locality; 25.0% in middle (area A) vs 14.0% in western parts (area B) (Table 1). Based on the fact that both areas belong to the South Sea and the distance between the areas is not so far (approximately 50 km), it seems that the locality where the anchovies were caught did not affect much difference in the prevalence. Rather, season (October in area A and May in area B) of fishery might have been a more important factor for the difference. Previous studies on various fish species have described seasonal variations in the prevalence of anisakid larvae, including *A. simplex*, *Con-*

tracaecum sp., and *H. aduncum* [32]. Further studies are needed to confirm seasonal tendencies in the prevalence of anisakid larvae in anchovies caught in the South Sea.

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CONFLICT OF INTEREST

We have no conflict of interest related to this work.

REFERENCES

1. Chai JY, Murrell KD, Lymbery AJ. Fish-borne parasitic zoonoses: status and issues. *Int J Parasitol* 2005; 35: 1233-1254.
2. van Thiel PH, Kuiper FC, Roskam RT. A nematode parasitic to herring, causing acute abdominal syndromes in man. *Trop Geogr Med* 1960; 2: 97-113.
3. Kim CH, Chung BS, Moon YI, Chun SH. A case report on human infection with *Anisakis* sp. in Korea. *Korean J Parasitol* 1971; 9: 39-43 (in Korean).
4. Sohn WM, Na BK, Kim TH, Park TJ. Anisakiasis: report of 15 gastric cases caused by *Anisakis* type I larvae and a brief review of Korean anisakiasis cases. *Korean J Parasitol* 2015; 53: 465-470.
5. Im KI, Shin HJ, Kim BH, Moon SI. Gastric anisakiasis cases in Cheju-do, Korea. *Korean J Parasitol* 1995; 33: 179-186 (in Korean).
6. Choi SC, Lee SY, Song HO, Ryu JS, Ahn MH. Parasitic infections based on 320 clinical samples submitted to Hanyang University, Korea (2004-2011). *Korean J Parasitol* 2014; 52: 215-220.
7. Noh JH, Kim B, Kim SM, Ock M, Park MI, Goo JY. A case of acute gastric anisakiasis provoking severe clinical problems by multiple infection. *Korean J Parasitol* 2003; 41: 97-100.
8. Lim H, Jung BK, Cho J, Yooyen T, Shin EH, Chai JY. Molecular diagnosis of cause of anisakiasis in humans, South Korea. *Emerg Infect Dis* 2015; 21: 342-344.
9. Song H, Jung BK, Cho J, Chang T, Huh S, Chai JY. Molecular identification of *Anisakis* larvae extracted by gastrointestinal endoscopy from health check-up patients in Korea. *Korean J Parasitol* 2019; 57: 207-211.
10. Sohn WM, Chai JY. Anisakiosis (Anisakidosis). In Palmer SR, Soulsby L, Torgerson PR, Brown WG, eds, *Oxford Textbook of Zoonoses-Biology, Clinical Practice, and Public Health Control*. 2nd ed. Oxford, UK. Oxford University Press. 2011, pp 774-786.
11. Song SB, Lee SR, Chung HH, Han NS. Infection status of anisakid larvae in anchovies purchased from local fishery market near southern and eastern sea in Korea. *Korean J Parasitol* 1995;

- 33: 95-99 (in Korean).
12. Kim WS, Jeon CH, Kim JH, Kim DH, Oh MJ. Current status of anisakid nematode larvae infection in marine fishes caught from the coastal area of Korea between 2010 and 2012. *J Fish Pathol* 2012; 25: 189-197.
 13. Bak TJ, Jeon CH, Kim JH. Occurrence of anisakid nematode larvae in chub mackerel (*Scomber japonicus*) caught off Korea. *Int J Food Microbiol* 2014; 191: 149-156.
 14. Jeon CH, Setyobudi E, Kim JH. Occurrence and molecular identification of anisakid nematodes isolated from Pacific cod (*Gadus macrocephalus*) caught off Korea. *Food Control* 2016; 69: 100-107.
 15. Cho J, Lim H, Jung BK, Shin EH, Chai JY. *Anisakis pegreffii* larvae in sea eels (*Astroconger myriaster*) from the South Sea, Republic of Korea. *Korean J Parasitol* 2015; 53: 349-353.
 16. D'Amelio S, Mathiopoulos KD, Santos CP, Pugachev ON, Webb SC, Picanço M, Paggi L. Genetic markers in ribosomal DNA for the identification of members of the genus *Anisakis* (Nematoda: ascaridoidea) defined by polymerase-chain-reaction-based restriction fragment length polymorphism. *Int J Parasitol* 2000; 30: 223-226.
 17. Campana-Rouget PY, Biocca E. Une Nouvelle Espèce D'*Anisakis* Chez un Phoque Mediterranean. *Ann de Parasitol* 1955; 30: 477-480.
 18. D'Amelio S, Mathiopoulos KD, Brandonisio O, Lucarelli G, Doronzo F, Paggi L. Diagnosis of a case of gastric anisakidosis by PCR-based restriction fragment length polymorphism analysis. *Parassitologia* 1999; 41: 591-593.
 19. Fumarola L, Monno R, Ierardi E, Rizzo G, Giannelli G, Lalle M, Pozio E. *Anisakis pegreffii* etiological agent of gastric infections in two Italian women. *Foodborne Pathog Dis* 2009; 6: 1157-1159.
 20. Mattiucci S, Paoletti M, Borrini F, Palumbo M, Palmieri RM, Gomes V, Casati A, Nascetti G. First molecular identification of the zoonotic parasite *Anisakis pegreffii* (Nematoda: Anisakidae) in a paraffin-embedded granuloma taken from a case of human intestinal anisakiasis in Italy. *BMC Infect Dis* 2011; 11: 1-6.
 21. Mattiucci S, Fazii P, Rosa AD, Paoletti M, Megna AS, Glielmo A, de Angelis M, Costa A, Meucci C, Calvaruso V, Sorrentini I, Palma G, Bruschi F, Nascetti G. Anisakiasis and gastroallergic reactions associated with *Anisakis pegreffii* infection, Italy. *Emerg Infect Dis* 2013; 19: 496-499.
 22. Umehara A, Kawakami Y, Araki J, Uchida A. Molecular identification of the etiological agent of the human anisakiasis in Japan. *Parasitol Int* 2007; 56: 211-215.
 23. Arizono N, Yamada M, Tegoshi T, Yoshikawa M. *Anisakis simplex* sensu stricto and *Anisakis pegreffii*: biological characteristics and pathogenetic potential in human anisakiasis. *Foodborne Pathog Dis* 2012; 9: 517-521.
 24. Bilska-Zajac E, Rózycki M, Chmurzyńska E, Karamon J, Sroka J, Kochanowski M, Kusyk P, Cencek T. Parasites of Anisakidae family - Geographical distribution and threat to human health. *J Agric Sci Technol A* 2015; 5: 146-152.
 25. Li L, An R, Zhang L. A new species of *Hysterothylacium* (Nematoda: Anisakidae) from marine fishes from Yellow Sea, China, with a key to the species of the genus *Hysterothylacium*. *Zootaxa* 2007; 1614: 43-52.
 26. Marcogliese DJ. Larval parasitic nematodes infecting marine crustaceans in eastern Canada. 3. *Hysterothylacium aduncum*. *Proc Helminthol Soc Wash* 1996; 63: 12-18.
 27. Køie M. Aspects of the life cycle and morphology of *Hysterothylacium aduncum* (Rudolphi, 1802) (Nematoda, Ascaridoidea, Anisakidae). *Can J Zool* 1993; 71: 1289-1296.
 28. Li L, Zhao JY, Chen HX, Ju HD, An M, Xu Z, Zhang LP. Survey for the presence of ascaridoid larvae in the cinnamon flounder *Pseudorhombus cinnamomeus* (Temminck & Schlegel) (Pleuronectiformes: Paralichthyidae). *Int J Food Microbiol* 241: 108-116.
 29. Navone GT, Sardella NH, Timi JT. Larvae and adults of *Hysterothylacium aduncum* (Rudolphi, 1802) (Nematoda: Anisakidae) in fishes and crustaceans in the southwest Atlantic. *Parasite* 1998; 5: 127-136.
 30. Yagi K, Nagasawa K, Ishikura H, Nakagawa A, Sato N, Kikuchi K, Ishikura H. Female worm *Hysterothylacium aduncum* excreted from human: A case report. *Jpn J Parasitol* 1996; 45: 12-23.
 31. González-Amores Y, Clavijo-Frutos E, Salas-Casanova C, Alcain-Martínez G. Direct parasitological diagnosis of infection with *Hysterothylacium aduncum* in a patient with epigastralgia. *Rev Esp Enferm Dig (Madrid)* 2015; 107: 699-700.
 32. Strømnes E, Andersen K. "Spring rise" of whaleworm (*Anisakis simplex*; Nematoda, Ascaridoidea) third-stage larvae in some fish species from Norwegian waters. *Parasitol Res* 2000; 86: 619-624.