



4-Hydroxybenzaldehyde Restricts the Intracellular Growth of *Toxoplasma gondii* by Inducing SIRT1-Mediated Autophagy in Macrophages

Jina Lee^{1,2}, Jae-Won Choi^{1,2}, Hye Young Han¹, Woo Sik Kim³, Ha-Yeon Song³, Eui-Baek Byun³, Eui-Hong Byun⁴, Young-Ha Lee¹, Jae-Min Yuk^{1,2,*}

¹Department of Infection Biology and Medical Science, College of Medicine, Chungnam National University, Daejeon 35015, Korea; ²Infection Control Convergence Research Center, College of Medicine, Chungnam National University, Daejeon 35015, Korea; ³Advanced Radiation Technology Institute, Korea Atomic Energy Research Institute, Jeongseup 56212, Korea; ⁴Department of Food Science and Technology, Kongju National University, Yesan 32439, Korea

Abstract: *Toxoplasma gondii* is an intracellular protozoan parasite that infects approximately one third of the human population worldwide. Considering the toxicity and side effects of anti-toxoplasma medications, it is important to develop effective drug alternatives with fewer and less severe off-target effects. In this study, we found that 4-hydroxybenzaldehyde (4-HBA) induced autophagy and the expression of NAD-dependent protein deacetylase sirtuin-1 (SIRT1) in primary murine bone marrow-derived macrophages (BMDMs). Interestingly, treatment of BMDMs with 4-HBA significantly reduced the number of macrophages infected with *T. gondii* and the proliferation of *T. gondii* in infected cells. This effect was impaired by pretreating the macrophages with 3-methyladenine or wortmannin (selective autophagy inhibitors) or with sirtinol or EX527 (SIRT1 inhibitors). Moreover, we found that pharmacological inhibition of SIRT1 prevented 4-HBA-mediated expression of LC3-phosphatidylethanolamine conjugate (LC3-II) and the colocalization of *T. gondii* parasitophorous vacuoles with autophagosomes in BMDMs. These data suggest that 4-HBA promotes antiparasitic host responses by activating SIRT1-mediated autophagy, and 4-HBA might be a promising therapeutic alternative for the treatment of toxoplasmosis.

Key words: *Toxoplasma gondii*, toxoplasmosis, autophagy, sirtinol, 4-hydroxybenzaldehyde, NAD-dependent protein deacetylase sirtuin-1, LC3-phosphatidylethanolamine conjugate

INTRODUCTION

Toxoplasma gondii, a member of the Apicomplexa phylum, is an obligate protozoan pathogen that can invade and replicate in almost all nucleated cells of warm-blooded animals. Almost a third of the global human population is infected with *T. gondii* [1,2]. For most immunocompetent individuals, infection is asymptomatic, chronic, and life-long; however, *T. gondii* infection in immunocompromised individuals and pregnant women can cause severe illness with high morbidity and mortality rates [2,3]. First-line treatment for toxoplasmosis is combined administration of pyrimethamine and sulfadiazine; however, because of their potential off-target effects, there is an

urgent need to develop therapeutic alternatives with fewer, more benign side effects [4].

Autophagy, which plays a pivotal role in maintaining cellular homeostasis, is a mechanism by which cells remove dysfunctional or dispensable cellular components such as damaged cytosolic organelles and long-lived/misfolded proteins through the fusion of autophagosomes and lysosomes [5,6]. Accumulating evidence suggests that selective autophagy, also known as xenophagy, helps protect the host against diverse infectious agents including bacteria, viruses, and protozoa [7,8]. During *T. gondii* infection, the host's autophagy machinery contributes to the restriction of intracellular survival by maintaining a balance between the host immune response and exploitation of the host by the parasite [9].

Prior studies of 4-hydroxybenzaldehyde (4-HBA), an essential bioactive constituent of the traditional Chinese herb *Gastrodiae Rhizoma*, reported anti-diabetic properties [10], antioxidant properties [11], and angiogenesis-promoting effects [12]. We previously reported that 4-HBA protects against DSS-in-

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*Corresponding author (yjaemin0@cnu.ac.kr)

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duced colitis by regulating immune tolerance and excessive inflammatory responses [13]. A recent study demonstrated that 4-HBA increased the susceptibility to amphenicol antibiotics including chloramphenicol and thiamphenicol in *Acinetobacter baumannii* [14]; however, the function of 4-HBA in infectious diseases has not yet been characterized.

In this study, we investigated the immunomodulatory properties of 4-HBA in *T. gondii*-infected murine bone marrow-derived macrophages (BMDMs). We found that 4-HBA restricts the intracellular growth of *T. gondii* via SIRT1-mediated autophagy activation. Collectively, our results suggest that 4-HBA may be a promising therapeutic alternative to treat patients infected with *T. gondii*.

MATERIALS AND METHODS

Mice and cell culture

Wild-type C57BL/6 mice were purchased from Koatech (Gyeonggi-do, Korea). Animal-related experimental procedures were approved by the Institutional Animal Care and Use Committee, Chungnam National University College of Medicine (CNU-00706; Daejeon, Republic of Korea). Bone marrow-derived macrophages were differentiated during 5–7 days in medium with macrophage colony-stimulating factor, as described previously [15]. The culture medium is consisted of Dulbecco's modified Eagle's medium (DMEM; Welgene) supplemented with 10% fetal bovine serum (FBS, Gibco BRL) and 1% Antibiotic-Antimycotic (Gibco™ Antibiotic-Antimycotic (100X); Gibco BRL). The human retinal pigment epithelial cell line ARPE-19 (American Type Culture Collection, Manassas, Virginia, USA) were grown in DMEM/F-12 (Welgene) with 10% FBS and 1% Antibiotic-Antimycotic.

Parasite preparation

T. gondii RH strain was multiplied in ARPE-19 cells at a multiplicity of infection (MOI) of 5 and grown for 2-3 days at 37°C and 5% CO₂. *T. gondii* RH strain expressing transgenic green fluorescent protein (GFP-RH) were kindly provided by Dr. Yoshifumi Nishikawa (Obihiro University of Agriculture and Veterinary Medicine, Japan). Host cell debris and parasites were washed in phosphate-buffered saline (PBS) after spontaneous host cell rupture. Final pellet was suspended in cold DMEM, and then passed through a 26-gauge needle and a 5.0 µm pore filter (Millipore, Billerica, Massachusetts, USA).

Reagents and antibodies

4-HBA (144088), 3-methyladenine (3-MA, M9281), EX-527 (E7034), sirtinol (S7942), wortmannin (WM, W1628), dimethylsulfoxide (DMSO, D2650) or LC3 (L8918) were from Sigma-Aldrich. β-Tubulin (ab6046) and SIRT1 (AB28170) were purchased from Abcam. Ethanol or DMSO was added to macrophages cultures at 0.05% (v/v) and used as a solvent control.

Cell viability assays

The cytotoxicity effects of 4-HBA on BMDMs were determined using cell count kit (CCK) 8 (Dojindo Molecular Technologies), according to the manufacturer's instructions. CCK8 solution (10 µl) was added after cells were incubated with 4-HBA for 18 or 48 hr. Absorbance was measured at 450 nm on a microplate reader (SpectraMax ABS Plus, Molecular Devices).

RNA extraction, real-time quantitative PCR, and western blot analysis

RNA extraction and real-time quantitative PCR were performed as described previously [15]. The sequences of the primers used were as follows: *Sag1* (forward: 5'-ATCGCCTGAGAAGCATCACT-3'; reverse: 5'-GCCAAAATGGAAACGTGACT-3'), β-actin (forward: 5'-TCATGAAGTGTGACGTTGACATCCGT-3'; reverse: 5'-CCTAGAAGCATTGCGGTGCACGATG-3').

Western blot analysis was performed as described previously [15]. Collected cell lysates was lysed using RIPA buffer (10 mM Tris-HCl at pH 8.0, 1 mM EDTA, 140 mM NaCl, 0.1% SDS, 0.1% sodium deoxycholate and 1% Triton X-100) containing a protease inhibitor cocktail (Roche). The protein extracts boiled with SDS sample buffer was subjected to sodium dodecyl sulfate polyacrylamide gel electrophoresis and then polyvinylidene fluoride membranes (Millipore Corp.). Chemiluminescence assay kit (ECL; Millipore Corp.) was used to develop membranes.

Quantification of intracellular *T. gondii*

BMDMs cultured on 22-mm glass coverslips were infected with GFP-RH strain for indicated time periods, and then coverslips were washed using warmed PBS. After fixation with 4% paraformaldehyde in PBS for 10 min and permeabilization with 0.25% Triton X-100 in PBS for 10 min, Texas Red®-X phalloidin (Life Technologies Corporation) and 4'6-diamidino-2-phenylindole (DAPI, Sigma) were used to visualize F-actin in the cytoskeleton and nucleus. Cover slides were analyzed by confocal laser scanning microscopy (Leica TCS SP8, Leica microsystems).

Immunofluorescence analyses

Immunofluorescence analysis of endogenous LC3 puncta and colocalization of LC3 with GFP-RH was performed as described previously [16]. Followed by fixation and permeabilization, cells in coverslips were stained with LC3 Ab (MBL International, PM036) for 2 hr at room temperature. After washing excess primary Ab with PBS, cells were incubated with Alexa Fluor 488-conjugated goat anti-rabbit IgG (Molecular Probes) or Alexa Fluor 594-conjugated goat anti-rabbit IgG (Molecular Probes) for 2 hr. Nucleus was stained with DAPI. Fluorescence images were obtained using confocal laser scanning microscopy. Each experiment for immunofluorescence microscopy contains at least 70 cells scored from 8 random fields for analysis of endogenous LC3 puncta. 70 internalized GFP-RH per experimental condition were counted for quantitative data of colocalization between LC3 with GFP-RH.

Statistical analysis

A 2-tailed Student's *t* test was used to analyze differences between independent experimental data (means \pm standard deviation [SD] or means \pm standard error [SEM]). Differences were deemed significant at *P*-value under 0.05.

RESULTS

Cytotoxic effects of 4-HBA in BMDMs

To evaluate the cytotoxicity of 4-HBA, we assessed the effects of 4-HBA on BMDM viability by CCK8 assay. Incubating BMDMs with 10 μ M 4-HBA had no significant effect on cell viability after 18 hr (Fig. 1A) or 48 hr (Fig. 1B); however, cell viability decreased by approximately 35% at 24 hr (Fig. 1A) and 92% at 48 hr after treatment with 50 μ M 4-HBA (Fig. 1B). Based on these

results, for subsequent experiments we used 10 μ M 4-HBA, a concentration that did not induce cytotoxicity in BMDMs, and evaluated the antiparasitic effects of 4-HBA on *T. gondii* infection.

4-HBA inhibited the intracellular growth of *T. gondii* in BMDMs

T. gondii can invade and live within all nucleated cells (including macrophages and dendritic cells) by forming parasitophorous vacuoles (PVs) [17], thus, we tested whether 4-HBA exhibit antiparasitic effects against *T. gondii* infection in BMDMs. BMDMs were infected with a GFP-expressing RH strain of *T. gondii* (GFP-RH) for the indicated periods (Fig. 2A-C), and then evaluated for intracellular growth of *T. gondii*. As shown in Fig. 2A and B, the number of *T. gondii*-infected cells was markedly suppressed by 4-HBA treatment in a dose-dependent manner. Moreover, intracellular proliferation of the parasite was also inhibited in 4-HBA-stimulated BMDMs when compared to BMDMs treated with SC (Fig. 2A and C). In addition, we assessed the antiparasitic effects of 4-HBA by analyzing the expression levels of *T. gondii* surface antigen 1 (*SAG1*). As shown in Fig. 2D, *SAG1* mRNA expression significantly decreased in 4-HBA-treated BMDMs in a concentration-dependent manner. These results indicate that 4-HBA plays an essential role in the activation of antiparasitic responses to *T. gondii* infection.

4-HBA-induced autophagy played an essential role in the activation of antiparasitic responses in primary murine macrophages

Previous studies demonstrated that autophagy contributes to host immune defenses against *T. gondii* infection [8,18]; however, the role of 4-HBA in autophagy activation has not been characterized. To determine whether 4-HBA treatment

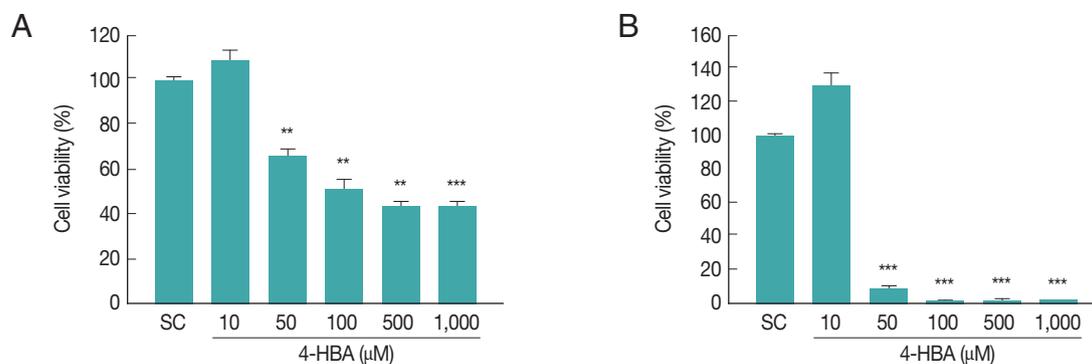


Fig. 1. Cytotoxic effect of 4-HBA on BMDMs by CCK8 assays. (A, B) Evaluation of cell viability at 18 hr (A) or 48 hr (B) after 4-HBA treatment in BMDMs. ***P* < 0.01, ****P* < 0.001. SC, solvent control.

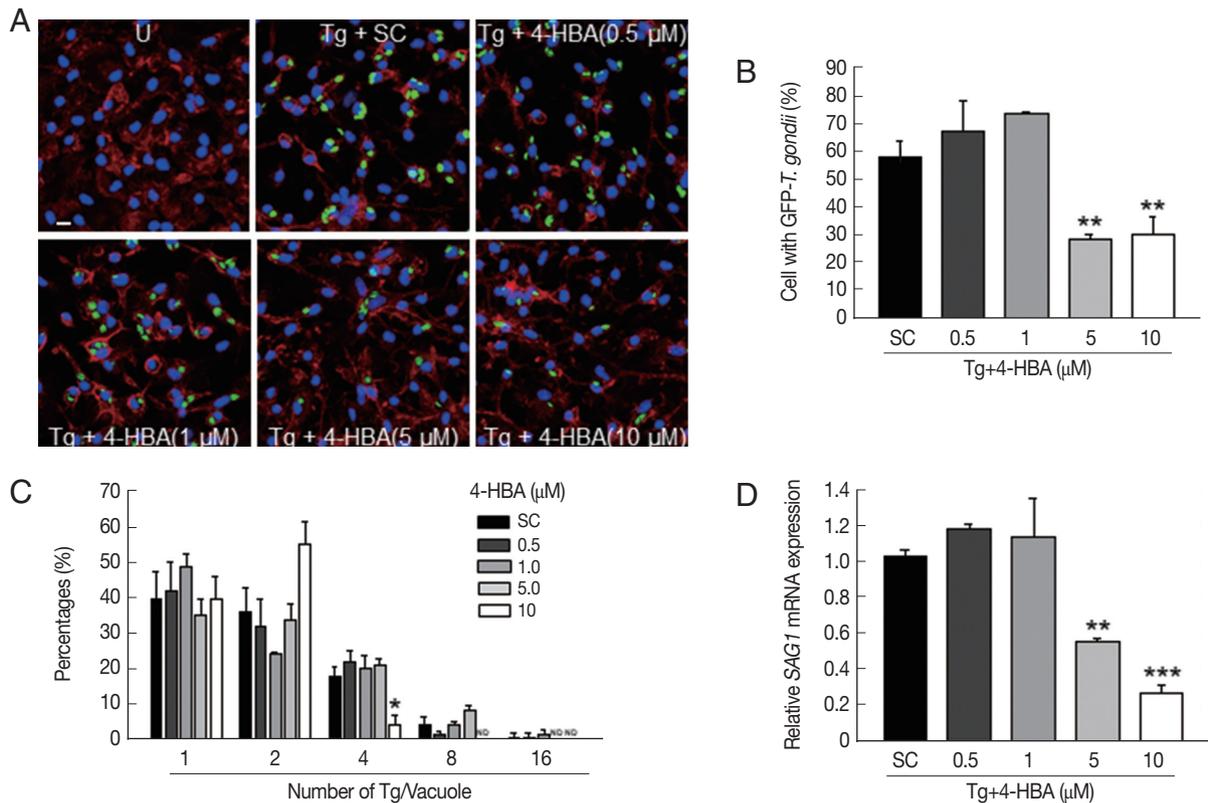


Fig. 2. 4-HBA increased antiparasitic activity in *T. gondii*-infected macrophages. (A-D) BMDMs were infected with GFP-RH strain (A, B, and C) or RH strain (D) for 2 hr (moi = 1) and then treated with 4-HBA for 18 hr. (A) Representative images showing *T. gondii* invasion into and growth in BMDMs. Scale bar = 10 μm. (B) Proportion of cells infected with *T. gondii* to total cells. (C) Number of parasites per vacuole. (D) Quantitative r-t PCR on SAG1 mRNA. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, U, untreated; SC, solvent control; Tg, *T. gondii*; ND, not detected.

induces autophagy in macrophages, we evaluated the number of cells with microtubule-associated protein 1 light chain 3 (LC3), an essential autophagy effector. As shown in Fig. 3A, 4-HBA treatment increased the number of LC3 aggregates in BMDMs. We next attempted to investigate the role of autophagy in 4-HBA-mediated anti-toxoplasma activity. As shown in Fig. 3B, the anti-toxoplasma activity of 4-HBA was significantly attenuated by treating *T. gondii*-infected BMDMs with autophagy inhibitors (3MA or WM). Our results suggest that autophagy is required for 4-HBA-induced activation of antiparasitic responses in primary macrophages.

4-HBA activated the induction of SIRT1, which contributes to the activation of antiparasitic responses in primary murine macrophages

Growing evidence suggests that SIRT1 is an essential regulator of diverse physiological conditions and activities such as cancer, metabolism, angiogenesis, and neuronal development

[19,20]. Moreover, recent studies demonstrate that SIRT1 can regulate innate and adaptive immune responses, and impaired SIRT1 activity is closely linked with the pathogenesis of autoimmune and inflammatory diseases [21]. We found that 4-HBA increased the expression of SIRT1 within 2 hr, and the increase was sustained for up to 18 hr in BMDMs (Fig. 4A). Next, we investigated if SIRT1 was required for 4-HBA-induced antiparasitic responses. SAG1 mRNA expression decreased significantly in 4-HBA-treated BMDMs (Fig. 4B and C); however, these effects disappeared in the presence of EX-527 (Fig. 4B) or sirtinol (Fig. 4C), selective SIRT1 inhibitors. These data suggest that SIRT1 controls 4-HBA-mediated growth inhibition of *T. gondii* in primary murine macrophages.

SIRT1 is required for 4-HBA-mediated activation of autophagy in BMDMs

Previous studies demonstrate that SIRT1 regulates autophagy activation through the deacetylation of autophagy-related

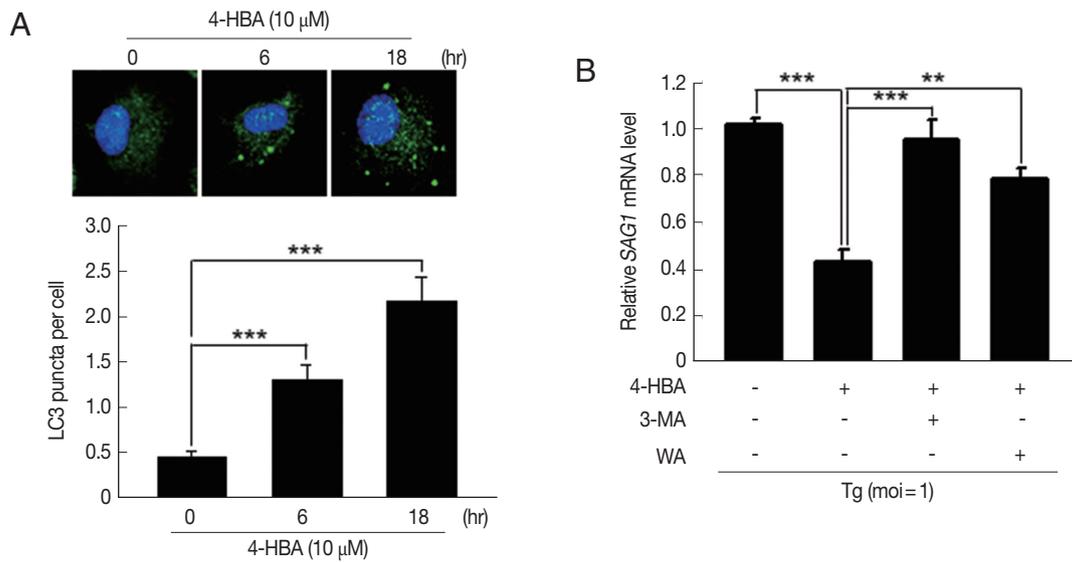


Fig. 3. 4-HBA-induced autophagy was required for the intracellular restriction of *T. gondii* in BMDMs. (A) Immunofluorescence microscopy on LC3 puncta formation in BMDMs treated with 4-HBA. Representative images (top; Scale bar = 10 μ m). LC3 punctate foci per cell (Lower panel). (B) SAG1 mRNA level quantified by rt-PCR. BMDMs were infected with *T. gondii* RH strain, treated with 3-MA (10 μ M) or WM (100 nM), and then incubated with 4-HBA (10 μ M) for 18 hr. ** P < 0.01, *** P < 0.001.

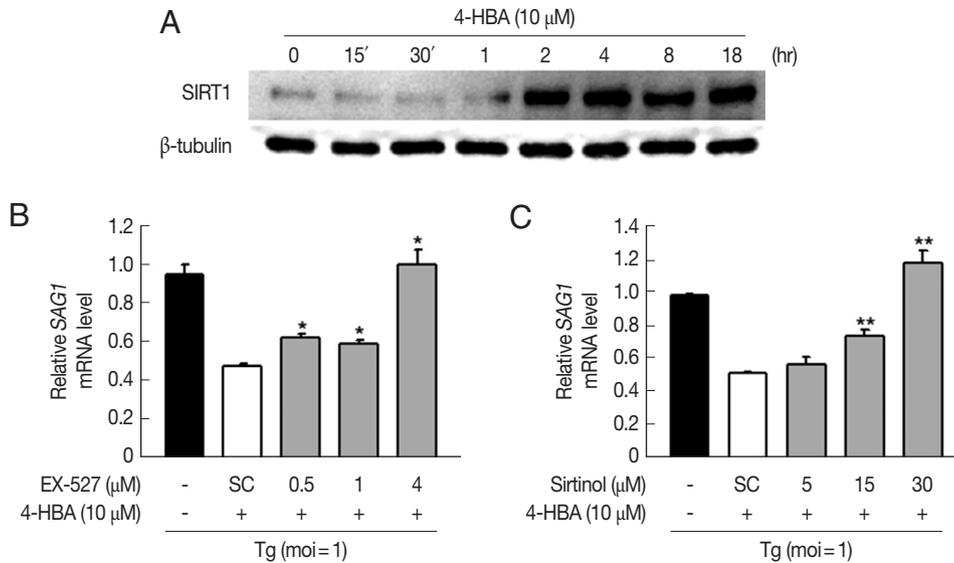


Fig. 4. Sirt1 was involved in 4-HBA-mediated activation of antiparasitic responses in BMDMs. (A) Immunoblot of SIRT1 and β -tubulin in BMDMs treated with 4-HBA. (B, C) SAG1 mRNA level. BMDMs were infected with RH strain in the presence of EX-527 (B) or sirtinol (C) and incubated with 4-HBA for 18 hr. * P < 0.05, ** P < 0.01. SC, solvent control; Tg, *T. gondii*.

genes (Atgs) and regulators of autophagy [22]. To examine whether 4-HBA-induced autophagy is mediated by SIRT1, we evaluated the formation of autophagosomes by BMDMs. We found that 4-HBA-mediated the increase of LC3-positive vesicles was significantly attenuated by pretreating BMDMs with EX-527 or sirtinol (Fig. 5A). Moreover, we also found that 4-HBA significantly increased the intracellular colocalization

of LC3-II-positive autophagic vacuoles with *T. gondii*-containing PVs in BMDMs as compared to non-treated infected cells; however, these colocalizations were significantly attenuated by EX-527 or sirtinol (Fig. 5B, C). These data suggest that SIRT1 is an essential mediator in 4-HBA-induced autophagy activation in primary macrophages.

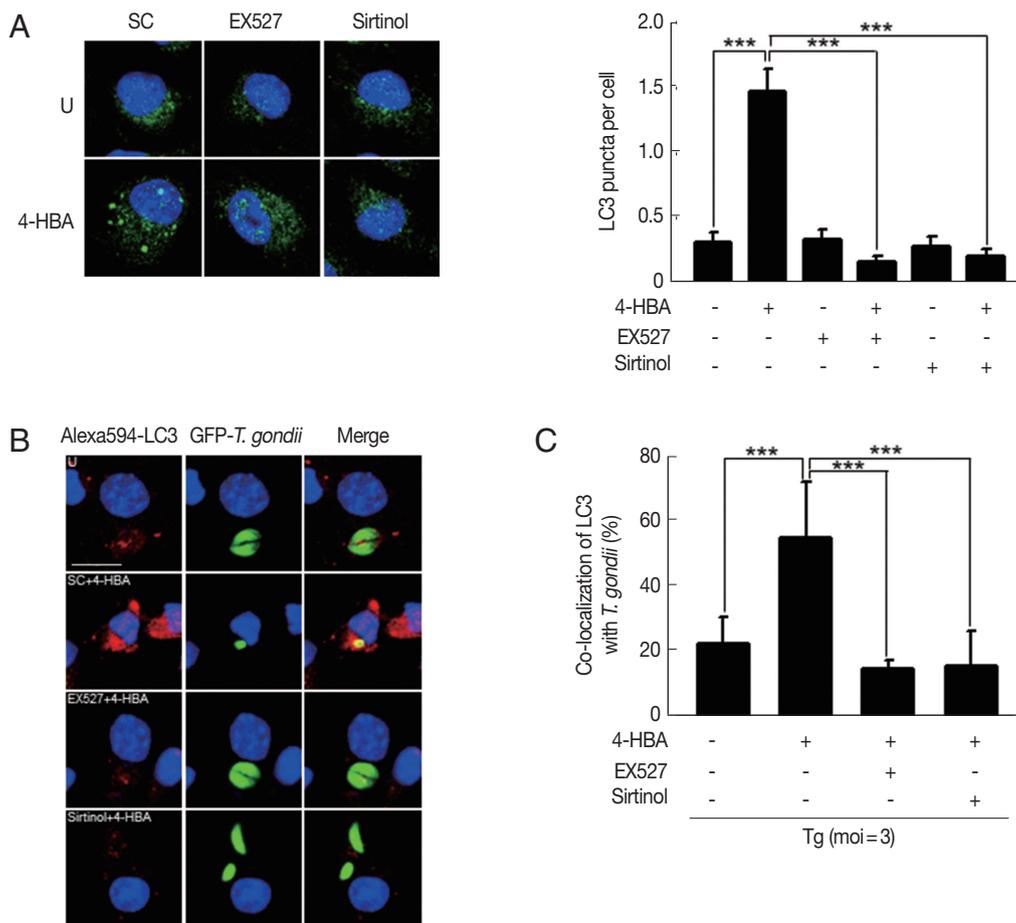


Fig. 5. Sirt1 contributed 4-HBA to induce formation and maturation of autophagosome and led activation of antiparasitic responses to *T. gondii* infection. (A) Immunofluorescence micrographs showing the formation of LC3 puncta in BMDMs treated with 4-HBA (10 μM; 18hr) in SC, EX-527 (4 μM), or sirtinol (30 μM). Left panel, representative images. Right panel, LC3 punctate foci in the cells. Scale bar= 10 μm. (B, C) BMDMs were infected with GFP-RH strain, followed by treatments of SC, EX-527 (4 μM), or sirtinol (30 μM), and then incubated with 4-HBA (10 μM) for 18 hr. Cells were stained with Alexa 594-conjugated LC3 (red) and DAPI (blue), (B) Immunofluorescence micrographs showing co-localization of GFP-RH strain (green) and LC3 (red). Scale bar= 10 μm. (C) Co-localization of LC3 with *T. gondii*. *****P*<0.001. SC, solvent control; Tg, *T. gondii*.

DISCUSSION

Autophagy is a tightly regulated homeostatic process that is essential to maintain cellular integrity, and impaired autophagy activation has been linked to the progression of physiological and pathological disorders [23]. Accumulating evidence suggests that xenophagy, a specialized, selective form of autophagy, is involved in activating protective host immunity responses via canonical and non-canonical pathways [24]; thereby, eliminating intracellular pathogens [7,25]. Many studies have evaluated flavonoids, a group of plant metabolites, as potential autophagy-inducing agents, which may be promising therapeutic alternatives for various autophagy de-

regulation-related diseases [26]. During *T. gondii* infection, autophagy can either protect the host or promote the pathophysiology of the parasite. Initial studies demonstrate that *T. gondii* promotes autophagy in host cells so they may utilize cellular nutrients in the early phases of infection [27]; however, in late infection phases, proliferating *T. gondii* suppress autophagy in host cells [28]. Recent studies suggest that canonical [29] or non-canonical autophagy [30,31] are involved in the restriction of intracellular *T. gondii*.

Protozoan parasites of the phylum Apicomplexa can survive and proliferate within diverse cell types. They are the etiological agent for various human diseases including toxoplasmosis, malaria, and cryptosporidiosis. 4-HBA is a hydroxybenzalde-

hyde and a major bioactive compound of *Gastrodiae Rhizoma*. A recent study suggests that 4-HBA, isolated from *Alpinia conchigera*, is one of the antimicrobials that affects *Staphylococcus aureus*, *Microsporum canis*, *Candida albicans*, and *Trichophyton rubrum* [32]. However, the roles of 4-HBA in autophagy and antiparasitic responses have not been investigated. In this study, we found that 4-HBA induced autophagy in primary macrophages, which is required to inhibit the intracellular growth of *T. gondii*.

To date, 7 mammalian Sirts have been identified. They are essential modulators of cell survival/death and energy status [33]. Among them, SIRT1, a class III histone deacetylase and NAD⁺-dependent protein, acts as a metabolic sensor of cellular energy. [19]. Several studies have demonstrated that SIRT1 activation is required for the regulation of various cellular responses [34]. For example, resveratrol, a SIRT1 activator, promotes SIRT1-dependent autophagy and ameliorates excessive inflammatory responses [35,36]. Moreover, we previously reported that the AMP-activated protein kinase pathway and SIRT1-induced orphan nuclear receptor estrogen-related receptor α (ESRRA) expression are required for the activation of autophagy and antimicrobial responses against *Mycobacterium tuberculosis* infection [37]. Herein, we found that the inactivation of SIRT1 inhibited 4-HBA-induced autophagy and antiparasitic responses, indicating that SIRT1-mediated autophagy is crucial for host-protective effects of 4-HBA against *T. gondii* infection.

In summary, we found that 4-HBA significantly restricted the intracellular growth of *T. gondii* via SIRT1-mediated autophagy in primary murine macrophages, and mediated *T. gondii* infection. These findings suggest that 4-HBA may be a promising, novel toxoplasmosis therapeutic. In future studies, we will evaluate the potential synergy between 4-HBA and standard therapeutics, such as pyrimethamine and sulfadiazine, to treat *T. gondii* infection.

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CONFLICT OF INTEREST

The authors have no financial conflict of interests.

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