



# Larval Gnathostomes and Zoonotic Trematode Metacercariae in Fish from a Local Market in Yangon City, Myanmar

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**Abstract:** A survey was performed to investigate the infection status of zoonotic helminth larvae in fish from a local market of North Dagon District in Yangon City, Myanmar. A total of 486 fish in 13 species were collected 8 times from December 2015 to December 2019. All fish were transported under ice to a laboratory in Korea and examined for helminth larvae using artificial digestion method. Larval gnathostomes and metacercariae of more than 8 zoonotic trematode species, i.e., *Opisthorchis viverrini*, *Haplorchis taichui*, *H. pumilio*, *H. yokogawai*, *Centrocestus* spp., *Stellantchasmus falcatus*, *Pygidioopsis cambodiensis*, and *Procerovum* sp., were detected. Larval gnathostomes were found in 58 (16.0%) out of 362 fish of 6 species, with mean intensity of 2.8 per fish infected. Metacercariae of *O. viverrini* were detected in 10 (2.9%) out of 349 fish of 5 species, with mean intensity of 16.9 per fish infected. Metacercarial prevalences of 4 intestinal flukes, *H. taichui*, *H. pumilio*, *H. yokogawai*, and *Centrocestus* spp., were 16.8%, 26.0%, 12.5%, and 15.0% in the positive fish species, respectively, and mean metacercarial intensity was 63.3, 26.8, 86.2, and 8.7 per fish infected. Metacercariae of *S. falcatus* and *P. cambodiensis* were detected only from the mullet, *Chelon macrolepis*. Metacercariae of *Procerovum* sp. were found in *Channa striata* and *Anabas testudineus*. Collectively, it was confirmed that the fish were infected with gnathostome larvae and metacercariae of *O. viverrini* and intestinal flukes in Yangon City, Myanmar.

**Key words:** *Gnathostoma* sp., *Opisthorchis viverrini*, *Haplorchis taichui*, *Haplorchis pumilio*, *Haplorchis yokogawai*, *Stellantchasmus falcatus*, *Pygidioopsis cambodiensis*, *Centrocestus* spp., *Procerovum* sp., zoonotic helminth larva

Fishborne-zoonotic helminths (FZH), including *Gnathostoma* spp. and trematodes, provoke a remarkable morbidity in residents of some Asian countries, i.e., Lao PDR, Vietnam, Cambodia, Thailand, the Philippines, Taiwan, China, and Korea. This helminth group causes serious economic damage in the industry of fish aquaculture [1-4]. FZH infections in humans are mainly localized in riverside areas, in which riparian residents are vulnerable to be infected by habitual consumption of raw and/or fermented fish containing infective larvae. Especially, the riverside areas in Mekong river basin in Vietnam, Lao PDR, Cambodia, and Thailand are known to be highly

endemic with FZH infections [5-9].

Epidemiological studies on helminthic infections have not been actively performed in the Republic of the Union of Myanmar (Myanmar). Most studies if any were surveys on soil-transmitted helminthiasis [10-14]. However, Aung et al. [15] and Sohn et al. [16] reported the endemic status of opisthorchiasis in Myanmar. Won et al. [17] molecularly diagnosed 2 cases of *Taenia saginata* infection in Yangon, Myanmar. Chai et al. [18] surveyed the infection status of zoonotic trematode metacercariae (ZTM) in fish from a local market in Yangon, and they described the morphological characteristics of adult flukes recovered from experimental animals. On the other hand, surveys on larval gnathostome infections in fish hosts were performed 3 times in Myanmar [19-21]. Chai et al. [19] examined 10 freshwater fish in 3 species, i.e., 6 catfish (*Parasilurus* sp.), 3 freshwater breams (*Tilapia* sp.), and 1 snakehead (*Channa* sp.), from Yangon, Myanmar. Jung et al. [20] exam-

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ined total 15 snakeheads (*Channa striata*) from a suburban area of Naypyidaw (the capital of Myanmar). Chai et al. [21] surveyed larval gnathostome infections in 37 Asian swamp eels (*Monopterus albus*) purchased from a local market in Yangon. These studies were performed with limited numbers of fish in limited fish species, and/or probable fish hosts of *Gnathostoma* spp. Therefore, in the present study, we examined total 486 fish in 13 species to reveal the infection status of zoonotic helminth larvae (ZHL), including larval gnathostomes and ZTM, in Yangon, Myanmar.

We purchased a total of 486 fish of 13 species in a local market in North Dagon District [14], Yangon City, Myanmar, 8 times (December 2015, August and November 2016, November 2017, June and December 2018, 2019). All collected fish were transported with ice to the laboratory of Department of Parasitology and Tropical Medicine, Gyeongsang National University College of Medicine, Jinju, Korea. The length and weight of fish were individually measured and identified the fish species with the aid of the FishBase website (<http://www.fishbase.org/search.php>) (Table 1) [22]. Individual fish was finely ground with a mortar and pestle, the ground fish meat was mixed with artificial gastric juice and the mixture was incubated at 36°C for about 2 hr. The digested material was filtered through a 5×5 mm<sup>2</sup> mesh, and washed with 0.85% saline until the supernatant became clear. The sediment was carefully examined under a stereomicroscope. Larval gnathostomes and ZTM were separately collected viewing from the general feature, and were counted to get hold of the prevalence (%) and intensity of infection (no. of larvae per fish infected) by fish species [18,21].

Larval gnathostomes were detected from 58 (16.0%) out of 362 fish of 6 species (46.2%), i.e., *Channa lucius*, *Anabas testudineus*, *Chelon macrolepis*, *Channa striata*, *Heteropneustes kemratensis*, and *Channa* sp., and their mean intensity was 2.8 per fish infected. The infection status of larval gnathostomes by fish species is presented in Table 2. The advanced third-stage larvae (AdL<sub>3</sub>) of gnathostomes detected (n=17) from forest snakeheads, *C. lucius*, and climbing perch, *A. testudineus* were

**Table 1.** Fish purchased in a local fish market in North Dagon District, Yangon City, Myanmar

Fish species	No. of fish collected <sup>a</sup>	Length (cm)		Weight (g)	
		Range	Average	Range	Average
<i>Channa lucius</i>	105	9.9-19.4	13.5	15.7-152.3	46.9
<i>Anabas testudineus</i>	100	7.3-14.7	11.4	12.7-115.0	52.5
<i>Thynnichthys thynnoides</i>	79	7.3-16.0	9.1	6.8-87.5	17.9
<i>Chelon macrolepis</i>	78	6.2-18.0	12.4	3.1-97.9	38.9
<i>Channa striata</i>	55	12.3-23.5	17.0	26.6-178.6	73.4
<i>Puntius aurotaeniatus</i>	15	4.6-9.2	5.2	2.3-8.2	3.4
<i>Heteropneustes kemratensis</i>	14	13.2-18.7	16.7	26.0-83.5	55.8
<i>Cirrhinus cirrhosis</i>	10	13.8-26.0	17.2	39.3-293.2	99.2
<i>Puntioplites</i> sp.	10	7.4-13.8	10.0	8.8-58.4	24.4
<i>Channa</i> sp.	10	13.5-18.6	16.2	39.3-92.6	70.8
<i>Hypsibarbus lagleri</i>	7	9.7-14.4	12.3	22.2-76.3	50.4
<i>Ctenopharyngodon idella</i>	2	8.1-10.0	9.1	9.0-16.3	12.7
<i>Esomus altus</i>	1	-	9.5	-	8.0

<sup>a</sup>A total of 486 fish in 13 species.

**Table 2.** Infection status of *Gnathostoma* sp. larvae by species of fish from Yangon City, Myanmar

Fish species	No. of fish examined	No. of fish infected (%)	No. of larvae detected	
			Range	Average
<i>Channa lucius</i>	105	24 (22.9)	1-18	4.5
<i>Anabas testudineus</i>	100	18 (18.0)	1-2	1.2
<i>Chelon macrolepis</i>	78	2 (2.6)	-	1.0
<i>Channa striata</i>	55	11 (20.0)	1-11	2.9
<i>Heteropneustes kemratensis</i>	14	1 (7.1)	-	1.0
<i>Channa</i> sp.	10	2 (20.0)	-	1.0
Total	362	58 (16.0)	1-18	2.8

2.05-3.90 (av. 2.911) mm long and 0.20-0.36 (av. 0.311) mm wide. They had a characteristic head bulb (av.  $0.077 \times 0.187$  mm) with 4 rows of hooklets, a muscular long esophagus (av. 0.955 mm long), 2 pairs of cervical sac (av. 0.580 mm long), and a short tail (av. 0.059 mm long). The mean number of hooklets was 40, 43, 46, and 48 on the 1st, 2nd, 3rd, and 4th row, respectively. They were more or less smaller than those (from swamp eels and Chinese edible frogs) of previous studies in Myanmar [21,23].

More than 8 species, i.e., *Opisthorchis viverrini*, *Haplorchis taichui*, *H. pumilio*, *H. yokogawai*, *Centrocestus* spp., *Stellantchasmus falcatus*, *Pygidioopsis cambodiensis*, and *Procerovum* sp., of ZTM

were detected. The metacercariae of *O. viverrini* were found in 10 (2.9%) out of 349 fish in 5 species (38.5%), i.e., *C. lucius*, *A. testudineus*, *Thynnichthys thynnoides*, *C. striata*, and *Puntioplites* sp. and their mean intensity was 16.9 per fish infected (Table 3). The metacercariae of *H. taichui* were detected from 45 (16.8%) out of 268 fish in 7 species (53.9%), and their mean intensity was 63.3 per fish infected (Table 4). The metacercariae of *H. pumilio* were found in 102 (26.0%) out of 393 fish in 10 species (76.9%), and their mean intensity was 26.8 per fish infected (Table 5). *H. yokogawai* metacercariae were detected in 13 (12.5%) out of 104 fish in 3 species (23.1%), and their mean intensity was 86.2 per fish infected. *Centrocestus* spp.

**Table 3.** Infection status of *Opisthorchis viverrini* metacercariae by species of fish from Yangon City, Myanmar

Fish species	No. of fish examined	No. of fish infected (%)	No. of metacercariae detected	
			Range	Average
<i>Channa lucius</i>	105	6 (5.7)	1-50	21.2
<i>Anabas testudineus</i>	100	1 (1.0)	-	1.0
<i>Thynnichthys thynnoides</i>	79	1 (1.3)	-	2.0
<i>Channa striata</i>	55	1 (1.8)	-	38.0
<i>Puntioplites</i> sp.	10	1 (10.0)	-	1.0
Total	349	10 (2.9)	1-50	16.9

**Table 4.** Infection status of *Haplorchis taichui* metacercariae by species of fish from Yangon City, Myanmar

Fish species	No. of fish examined	No. of fish infected (%)	No. of metacercariae detected	
			Range	Average
<i>Channa lucius</i>	105	3 (1.3)	2-6	4.3
<i>Thynnichthys thynnoides</i>	79	17 (21.5)	1-61	7.9
<i>Channa striata</i>	55	3 (5.5)	2-36	17.0
<i>Puntioplites</i> sp.	10	8 (80.0)	1-728	163.5
<i>Hypsibarbus lagleri</i>	7	2 (28.6)	1-3	2.0
<i>Ctenopharyngodon idella</i>	2	2 (100)	6-10	8.0
Total	268	45 (16.8)	1-728	63.3

**Table 5.** Infection status of *Haplorchis pumilio* metacercariae by species of fish from Yangon City, Myanmar

Fish species	No. of fish examined	No. of fish infected (%)	No. of metacercariae detected	
			Range	Average
<i>Channa lucius</i>	105	23 (21.9)	1-890	94.9
<i>Anabas testudineus</i>	100	18 (18.0)	1-17	6.2
<i>Thynnichthys thynnoides</i>	79	30 (38.0)	1-12	2.5
<i>Channa striata</i>	55	9 (16.4)	1-20	6.6
<i>Puntius aurotaeniatus</i>	15	1 (6.7)	-	12.0
<i>Channa</i> sp.	10	2 (20.0)	1-92	46.5
<i>Puntioplites</i> sp.	10	8 (80.0)	1-728	3.1
<i>Cirrhinus cirrhosis</i>	10	4 (40.0)	-	4.0
<i>Hypsibarbus lagleri</i>	7	6 (85.7)	1-3	27.0
<i>Ctenopharyngodon idella</i>	2	1 (50.0)	-	1.0
Total	393	102 (26.0)	1-890	26.8

metacercariae were found in 54 (15.0%) out of 359 fish in 6 species (46.2%), and their mean intensity was 8.7 per fish infected. The infection status of *H. yokogawai* and *Centrocestus* spp. metacercariae by fish species are presented in detail in Tables 6 and 7. The metacercariae of *S. falcatus* and *P. cambodiensis* were detected in 8 (10.3%) and 11 (14.1%) out of 78 mullet, *C. macrolepis*, examined, and their mean intensities were 9.9 per fish infected each. *Procerovum* sp. metacercariae were found in 4 (7.3%) striped snakehead, *C. striata*, and 1 (1.0%) climbing perch, *A. testudineus*, and their mean intensities were 44.0 and 21.0 per fish infected.

In the present study, more than 9 species, i.e., *Gnathostoma* sp., *O. viverrini*, *H. taichui*, *H. pumilio*, *H. yokogawai*, *S. falcatus*, *P. cambodiensis*, *Procerovum* sp., and *Centrocestus* spp., including *C. formosanus*, of ZHL were detected in fish from a local market of North Dagon District in Yangon City, Myanmar. The presence of these ZHL in fish hosts were already reported in Myanmar [18-21]. However, large-scale surveys with lots of fish samples had not been performed in Myanmar. Therefore, by the present study, it has been reconfirmed that more than 9 etiologic agents of fishborne zoonotic helminthiasis are distributed in Yangon, Myanmar. Their infection status, i.e., prevalence and intensity of infection, was clarified in the aspect of host-parasite relationship.

Gnathostomiasis caused by larval *Gnathostoma* species is clinically characterized by creeping eruption in subcutaneous and intermuscular tissues due to migrating larvae. The larval

gnathostomes also occasionally invade the visceral organs, including livers and lungs, eyes, and even the central nerve system including brain [4]. Human cases of gnathostomiasis have occurred sporadically in Myanmar. More than 42 cutaneous cases and over 10 ocular cases have been reported since the first case in 1960 [24-28]. The infection status with larval gnathostomes were investigated in fish and frog hosts 4 times in Myanmar [19-21,23]. Chai et al. [19] found 2 AdL<sub>3</sub> of *G. spinigerum* from 2 catfish in the examination of 10 freshwater fish in 3 species, i.e., *Parasilurus* sp., *Tilapia* sp., and *Channa* sp., from Yangon. Jung et al. [20] also detected 2 AdL<sub>3</sub> of *G. spinigerum* in 2 snakeheads, *C. striata*, from a suburban area of Naypyidaw. Chai et al. [21] detected a total of 401 *G. spinigerum* AdL<sub>3</sub> in 33 (89.2%) out of 37 Asian swamp eels purchased in a local market of Yangon. Recently, Chai et al. [23] also detected 158 larval gnathostomes in 15 (75.0%) out of 20 Chinese edible frogs, *Hoplobatrachus rugulosus*, from a local market of Yangon. In this study, larval gnathostomes were found in 58 (16.0%) out of 362 fish in 6 species, i.e., *C. lucius*, *C. striata*, *A. testudineus*, *C. macrolepis*, *H. kemratensis*, and unidentified snakehead, and the forest snakehead, *C. lucius*, was the most prevalent fish host. Thus, 8 fish species, including catfish and swamp eel, already reported, and Chinese edible frogs are suggested to be the potential risk of human gnathostomiasis in Myanmar.

The endemicity of *O. viverrini* metacercariae (prevalence below 3.0%; intensity of infection about 17 per fish infected)

**Table 6.** Infection status of *Haplorchis yokogawai* metacercariae by species of fish from Yangon City, Myanmar

Fish species	No. of fish examined	No. of fish infected (%)	No. of metacercariae detected	
			Range	Average
<i>Thynnichthys thynnoides</i>	79	2 (2.5)	26-280	153.0
<i>Puntius aurotaeniatus</i>	15	3 (20.0)	1-385	129.3
<i>Puntioplites</i> sp.	10	8 (80.0)	5-218	53.3
Total	104	13 (12.5)	1-385	86.2

**Table 7.** Infection status of *Centrocestus* spp. metacercariae by species of fish from Yangon City, Myanmar

Fish species	No. of fish examined	No. of fish infected (%)	No. of metacercariae detected	
			Range	Average
<i>Channa lucius</i>	105	14 (13.3)	1-31	6.4
<i>Anabas testudineus</i>	100	14 (14.0)	2-58	19.7
<i>Thynnichthys thynnoides</i>	79	16 (20.3)	1-5	2.1
<i>Channa striata</i>	55	4 (7.3)	2-41	12.5
<i>Channa</i> sp.	10	2 (20.0)	1-11	6.0
<i>Puntioplites</i> sp.	10	4 (40.0)	1-3	2.0
Total	359	54 (15.0)	1-58	8.7

was very low like in Sohn et al. [16]. Among 5 positive fish species, 4, i.e., *C. lucius*, *C. striata*, *A. testudineus*, and unspecified *Puntioplites* sp., were previously reported [16], and only 1 species, *T. thynnoides*, is newly added to the second intermediate hosts of *O. viverrini* in Myanmar. The low-grade endemicity for opisthorchiasis in Myanmar was also confirmed by fecal examinations of people [15,16]. Aung et al. [15] found small trematode eggs in 34 (9.3%) out of 364 fecal samples of residents, and molecularly confirmed 18 cases to be opisthorchiasis infections. Sohn et al. [16] also detected small trematode eggs from 14 (0.7%) out of 2,057 fecal samples in 3 regions in Yangon. Accordingly, it is suggested that opisthorchiasis is maintained in low grade in the second intermediate fish host as well as in the definitive human host in Myanmar.

In the present study, *H. taichui* metacercariae were detected from 45 (16.8%) out of 268 fish in 7 (53.8%) species, i.e., *C. lucius*, *C. striata*, *T. thynnoides*, *C. cirrhosis*, *H. lagleri*, *C. idella*, and unspecified *Puntioplites* sp., and their mean intensity was 63.3 per fish infected. The prevalence was lower than that (42.3%) of Chai et al. [18], but the intensity of infection was more or less higher than that (42.9) of a previous study performed in Yangon, Myanmar [18]. Especially, the prevalences and intensities were relatively high in 2 fish species, *C. cirrhosis*, and *Puntioplites* sp. By this study and Chai et al. [18], a total of 11 fish species, i.e., *C. lucius*, *C. striata*, *T. thynnoides*, *C. cirrhosis*, *H. lagleri*, *C. idella*, *Puntius aurotaeniatus*, *Esomus altus*, *Puntioplites* sp., *Mystacoleucus* sp., and *Labeo* sp., are recorded as the second intermediate hosts of *H. taichui* in Myanmar.

The metacercariae of *H. pumilio* were detected from 102 (26.0%) out of 393 fish in 10 (76.9%) species, i.e., *C. lucius*, *C. striata*, *A. testudineus*, *T. thynnoides*, *C. cirrhosis*, *H. lagleri*, *C. idella*, *P. aurotaeniatus*, *Puntioplites* sp., and *Channa* sp., and their mean intensity was 26.8 per fish infected. The prevalence and intensity of infection were relatively low like in a previous study [18]. Based on the findings of present and previous studies, total 15 fish species, i.e., *C. lucius*, *C. striata*, *A. testudineus*, *T. thynnoides*, *C. cirrhosis*, *H. lagleri*, *C. idella*, *P. aurotaeniatus*, *Trichogaster pectoralis*, *E. altus*, *Puntioplites* sp., *Channa* sp., *Rhynogobius* sp., *Mystacoleucus* sp., and *Labeo* sp., are the second intermediate hosts of *H. pumilio* in Myanmar. In Chai et al. [18], *H. yokogawai* metacercariae were found in 5 fish species, i.e., *P. aurotaeniatus*, *E. altus*, *T. pectoralis*, *Mystacoleucus* sp., and *Labeo* sp., from Yangon, Myanmar, and they were heavily infected in 2 fish species, *P. aurotaeniatus* and *Mystacoleucus* sp. However, *H. yokogawai* metacercariae were detected in 3 fish species, *Puntio-*

*plites* sp., *P. aurotaeniatus*, and *T. thynnoides*, in this study. Therefore, total 7 fish species, i.e., *P. aurotaeniatus*, *E. altus*, *T. pectoralis*, *T. thynnoides*, *Puntioplites* sp., *Mystacoleucus* sp., and *Labeo* sp., have been found to be the second intermediate hosts of *H. yokogawai* in Myanmar.

*Centrocestus* spp. metacercariae, including *C. formosanus*, were found in 6 (46.2%) fish species, i.e., *C. lucius*, *C. striata*, *A. testudineus*, *T. thynnoides*, *Puntioplites* sp., and *Channa* sp., and their mean intensity was 8.7 per fish infected. The prevalence (15.0%) and intensity of infection were lower than those (50.8% and 25.8) of Chai et al. [18]. By the present study, 3 fish species, i.e., *C. lucius*, *Puntioplites* sp., and *Channa* sp., are newly added in the second intermediate hosts of *Centrocestus* spp. in Myanmar. As the second intermediate hosts of *Centrocestus* spp., a total of 11 fish species, i.e., *C. lucius*, *T. thynnoides*, *P. aurotaeniatus*, *E. altus*, *C. striata*, *A. testudineus*, *T. pectoralis*, *Puntioplites* sp., *Channa* sp., *Mystacoleucus* sp., and *Labeo* sp., have been listed in Myanmar.

In the present study, *Procerovum* sp. metacercariae were detected from 2 fish species, *C. striata* and *A. testudineus*, examined in August 2016. The climbing perch, *A. testudineus*, was previously reported as the second intermediate host of *Procerovum* spp., including *P. varium* in Vietnam, Lao PDR, and Myanmar [9,18,29,30]. The metacercariae of *S. falcatus* and *P. cambodiensis* were detected only in mullets, *Chelon macrolepis*, like in previous studies, including Chai et al. [18,31,32]. Chai et al. [18] detected a total of 36 *S. falcatus* metacercariae in 15 (34.1%) out of 44 mullets examined and they also found 38 metacercariae of *P. cambodiensis* in only 1 (2.2%) mullet from Yangon, Myanmar. In the present study, the prevalences were 10.3% and 14.1% and intensities of infection of 2 heterophyid flukes were commonly 9.9 per fish infected.

Conclusively, it has been confirmed that larval gnathostomes and more than 8 species of ZTM, i.e., *O. viverrini*, *H. taichui*, *H. pumilio*, *H. yokogawai*, *S. falcatus*, *P. cambodiensis*, *Procerovum* sp., and *Centrocestus* spp., including *C. formosanus*, were infected in fish from Yangon, Myanmar, and they can act as the etiologic agents of fishborne zoonotic helminthiasis although their endemicities were not so high. To reveal the detailed epidemiological situation of FZH infections in Myanmar, further studies on the intermediate and definitive hosts (humans and reservoir host animals) of FZH should be performed in the near future.



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## CONFLICT OF INTEREST

The authors don't have any conflicts of interest to declare.

## REFERENCES

- Chai JY, Murrell KD, Lymbery AJ. Fish-borne parasitic zoonoses: status and issues. *Int J Parasitol* 2005; 35: 1233-1254. <https://doi.org/10.1016/j.ijpara.2005.07.013>
- Chai JY. Intestinal flukes. In Murrell KD, Fried B eds, *Food-borne parasitic zoonoses*. New York, USA: Springer, 2007, pp 53-115.
- Hung NM, Madsen H, Fried B. Global status of fish-borne zoonotic trematodiasis in humans. *Acta Parasitol* 2013; 58: 231-258. <https://doi.org/10.2478/s11686-013-0155-5>
- Herman JS, Chiodini PL. Gnathostomiasis, another emerging imported disease. *Clin Microbiol Rev* 2009; 22: 484-492. <https://doi.org/10.1128/CMR.00003-09>
- Radomyos B, Wongsaroy T, Wilairatana P, Radomyos P, Praevanich R, Meesomboon V, Jongsuksuntikul P. Opisthorchiasis and intestinal fluke infections in northern Thailand. *Southeast Asian J Trop Med Public Health* 1998; 29: 123-127.
- Chai JY, Park JH, Han ET, Guk SM, Shin EH, Lin A, Kim JL, Sohn WM, Yong TS, Eom KS, Min DY, Hoang EH, Phommasack B, Insisienmay B, Rim HJ. Mixed infections with *Opisthorchis viverrini* and intestinal flukes in residents of Vientiane Municipality and Saravane Province in Laos. *J Helminthol* 2005; 79: 283-289. <https://doi.org/10.1079/joh.2005302>
- Chai JY, Han ET, Shin EH, Sohn WM, Yong TS, Eom KS, Min DY, Um JY, Park MS, Hoang EH, Phommasack B, Insisienmay B, Lee SH, Rim HJ. High prevalence of *Haplorchis taichui*, *Prosthodendrium molenkampi*, and other helminth infections among people in Khammouane Province, Lao PDR. *Korean J Parasitol* 2009; 47: 243-247. <https://doi.org/10.3347/kjp.2009.47.3.243>
- Dung DT, De NV, Waikagul J, Dalsgaard A, Chai JY, Sohn WM, Murrell KD. Fishborne intestinal zoonotic trematodiasis, Vietnam. *Emerg Infect Dis* 2007; 13: 1828-1833. <https://doi.org/10.3201/eid1312.070554>
- Phan VT, Ersbøll AK, Nguyen KV, Madsen H, Dalsgaard A. Farm-level risk factors for fishborne zoonotic trematode infection in integrated small-scale fish farms in northern Vietnam. *PLoS Negl Trop Dis* 2010; 4: 742-750. <https://doi.org/10.1371/journal.pntd.0000742>
- Nuchprayoon S, Sanprasert V, Kaewzaithim S, Saksirisampant W. Screening for intestinal parasitic infections among Myanmar migrant workers in Thai food industry: a high-risk transmission. *J Immigrant Minority Health* 2009; 11: 115-121. <https://doi.org/10.1007/s10903-008-9169-8>
- Htwe KT, Min TT, Mar TT, Lwin MM, Maw WW, Win MM. Detection of intestinal parasitic infestation among primary school children, Magway. *Myanmar Health Sci Res J* 2010; 22: 89-94.
- Aung T, Myat SM, Gabrielli AF, Montresor A. Control of soil-transmitted helminthiasis in Myanmar: Result of 7 years of deworming. *Trop Med Int Health* 2013; 18: 1017-1020. <https://doi.org/10.1111/tmi.12130>
- Htoon TT, Tun T, Oo KY, Thein W, Tin HH, Chai JY, Yong TS, Sohn WM. Status of infection with soil-transmitted helminths among primary school children in three selected townships of Yangon region. *Myanmar Health Sci Res J* 2015; 27: 221-226.
- Chai JY, Sohn WM, Hong SJ, Jung BK, Hong S, Cho S, Park JB, Kim IS, Kim S, Lee KH, Jeoung HG, Htoon TT, Tin HH. Effect of mass drug administration with a single dose of albendazole on *Ascaris lumbricoides* and *Trichuris trichiura* infection among schoolchildren in Yangon region, Myanmar. *Korean J Parasitol* 2020; 58: 195-200. <https://doi.org/10.3347/kjp.2020.58.2.195>
- Aung WPP, Htoon TT, Tin HH, Thinn KK, Sanpool O, Jongthawin J, Sadaow L, Phosuk I, Rodpai R, Intapan PM, Maleewong W. First report and molecular identification of *Opisthorchis viverrini* infection in human communities from lower Myanmar. *PLoS One*, 2017; 12: e0177130. <https://doi.org/10.1371/journal.pone.0177130>
- Sohn WM, Jung BK, Hong SJ, Lee KH, Park JB, Kim HS, Cho S, Htoon TT, Tin HH, Chai JY. Low-grade endemicity of opisthorchiasis, Yangon, Myanmar. *Emerg Infect Dis* 2019; 25: 1435-1437. <https://doi.org/10.3201/eid2507.190495>
- Won EJ, Jung BK, Song HM, Kim MS, Kim HS, Lee KH, Kim MJ, Shin MG, Shin JH, Suh SP, Hong SJ, Sohn WM, Htoon TT, Tin HH, Chai JY. Molecular diagnosis of *Taenia saginata* tapeworm infection in 2 schoolchildren, Myanmar. *Emerg Infect Dis* 2018; 24: 1156-1158. <https://doi.org/10.3201/eid2406.180217>
- Chai JY, Sohn WM, Na BK, Park JB, Jeoung HG, Hoang EH, Htoon TT, Tin HH. Zoonotic trematode metacercariae in fish from Yangon, Myanmar and their adults recovered from experimental animals. *Korean J Parasitol* 2017; 55: 631-641. <https://doi.org/10.3347/kjp.2017.55.6.631>
- Chai JY, Han ET, Shin EH, Park JH, Chu JP, Hirota M, Nakamura E, Nawa Y. An outbreak of gnathostomiasis among Korean emigrants in Myanmar. *Am J Trop Med Hyg* 2003; 69: 67-73.
- Jung BK, Lee JJ, Pyo KH, Kim HJ, Jeong HG, Yoon CH, Lee SH, Shin EH, Chai JY. Detection of *Gnathostoma spinigerum* third-stage larvae in snakeheads purchased from a central part of Myanmar. *Korean J Parasitol* 2008; 46: 285-288. <https://doi.org/10.3347/kjp.2008.46.4.285>

21. Chai JY, Sohn WM, Na BK, Park JB, Jeoung HG, Hoang EH, Htoon TT, Tin HH. Larval *Gnathostoma spinigerum* detected in Asian swamp eels, *Monopterus albus*, purchased from a local market in Yangon, Myanmar. *Korean J Parasitol* 2015; 53: 619-625. <https://doi.org/10.3347/kjp.2015.53.5.619>
22. Search FishBase [Internet]; Available from: <http://www.fishbase.org/search.php>
23. Chai JY, Jung BK, Lee KH, Ryu JY, Kim HS, Hong SJ, Htoon TT, Tin HH, Na BK, Sohn WM. Larval gnathostomes and spargana in Chinese edible frogs, *Hoplobatrachus rugulosus*, from Myanmar: potential risk of human infection. *Korean J Parasitol* 2020; 58: 467-473. <https://doi.org/10.3347/kjp.2020.58.4.467>
24. Gyi K. Intra-ocular gnathostomiasis. *Br J Ophthalmol* 1960; 44: 42-45. <https://doi.org/10.1136/bjo.44.1.42>
25. Khin T. Intra-ocular gnathostomiasis. *Br J Ophthalmol* 1968; 52: 57-60. <https://doi.org/10.1136/bjo.52.1.57>
26. Nomura Y, Nagakura K, Kagei N, Tsutsumi Y, Araki K, Sugawara M. Gnathostomiasis possibly caused by *Gnathostoma malaysiae*. *Tokai J Exp Clin Med* 2000; 25: 1-6.
27. Develoux M, Dekumyoy P, Baygon E, Aractingi S. Imported gnathostomiasis acquired in Myanmar. *Med Mal Infect* 2006; 36: 340-342 (in French). <https://doi.org/10.1016/j.medmal.2006.01.011>
28. Wai AP, Maw WW, Moe AC, Boonmars T, Nawa Y. Human gnathostomiasis in Myanmar: a review of local literature. *Southeast Asian J Trop Med Public Health* 2018; 49: 543-548.
29. Chai JY, De NV, Sohn WM. Foodborne trematode metacercariae in fish from northern Vietnam and their adults recovered from experimental hamsters. *Korean J Parasitol* 2012; 50: 317-325. <https://doi.org/10.3347/kjp.2012.50.4.317>
30. Eom KS, Park HS, Lee D, Sohn WM, Yong TS, Chai JY, Min DY, Rim HJ, Insisiengmay B, Phommasack B. Infection status of zoonotic trematode metacercariae in fishes from Vientiane Municipality and Champasak Province in Lao PDR. *Korean J Parasitol* 2015; 53: 447-453. <https://doi.org/10.3347/kjp.2015.53.4.447>
31. Chai JY, Sohn WM, Na BK, Jeoung HG, Sinuon M, Socheat D. *Stellantchasmus falcatus* (Digenea: Heterophyidae) in Cambodia: discovery of metacercariae in mullets and recovery of adult flukes in an experimental hamster. *Korean J Parasitol* 2016; 54: 537-541. <https://doi.org/10.3347/kjp.2016.54.4.537>
32. Sohn WM, Kim DC, Jung BK, Cho J, Chai JY. *Pygidiopsis cambodiensis* n. sp. (Digenea: Heterophyidae) from experimental hamsters infected with metacercariae in mullets from Cambodia. *Parasitol Res* 2016; 115: 123-130. <https://doi.org/10.1007/s00436-015-4727-1>

